



STUDIES ON THE ROLE OF SYNERGISTS FOR COUNTERING DEVELOPMENT OF INSECTICIDE RESISTANCE IN LARVAL POPULATION *Maruca vitrata* (Geyer) FROM MAJOR BLACKGRAM GROWING AREAS OF ANDHRA PRADESH

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ABSTRACT

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Role of synergist for countering the insecticide resistance mechanism was studied in both baseline and field populations during *rabi* 2018-19 at Insectary, Department of Entomology, S.V. Agricultural College, Tirupati. Higher per cent mortality was observed in larvae treated with insecticide + synergist against insecticide alone. Mean per cent mortality of insecticide + synergist and insecticide alone significantly differed. The highest percent mortality was observed with chlorantraniliprole alone and along with synergist followed by spinosad, thiodicarb, dichlorvos and chlorpyrifos alone and along with synergist respectively. In baseline population, the highest percent mortality was observed in insecticide and insecticide along with synergist followed by Prakasam, Nellore, Chittoor, Kurnool and Guntur districts larval population.

KEYWORDS: *Maruca vitrata*, chlorantraniliprole, spinosad, thiodicarb, dichlorvos, chlorpyrifos, PBO (Pyperonyl butoxide), sesamin.

INTRODUCTION

India is the world's largest producer of blackgram contributing 28 per cent to the global pulse basket from an area of about 37 per cent, as well as largest consumer. Blackgram is the 3rd important crop and it was cultivated over an area of 5.44 million hectares (*kharif* + *rabi*) and recorded a production of 3.56 million tonnes at a productivity level of 655 kg ha⁻¹. More than 90 per cent of urdbean production comes from nine states *viz.*, Madhya Pradesh, Rajasthan, Uttar Pradesh, Andhra Pradesh, Tamil Nadu, Maharashtra, Jharkhand, Gujarat and West Bengal. In Andhra Pradesh blackgram is cultivated in an area of 3.81 lakh ha and with a total production of 3.13 lakh tonnes with productivity of 946 kg ha⁻¹. Andhra Pradesh contributes 7.57 per cent and 9.53 per cent in area and production of blackgram in India (Indiastat, 2018). Blackgram contains 24 per cent protein, 3.2 per cent minerals, 59.6 per cent carbohydrate, amino acids and vitamins. It also contains 154 mg calcium, 9.1 mg iron and 38 mg β -carotene per 100g of split daul (Nene, 2006).

Insect pests are serious limiting factors in pulse production leading to reduced productivity. Blackgram is attacked by 40 to 60 insect species at different stages of the crop growth (Khazuria *et al.*, 2015). On an average, 2.5 to 3.0 million tonnes of pulses are lost annually due

to pest problems in India (Ramanujan, 2004). The yield losses on urd bean due to insect pests *Maruca vitrata* (Geyer), *Spodoptera litura* (Fabricius), thrips and pod bugs *etc.*, at various stages of the crop growth accounts 30 to 54.3 per cent in India (Dhuri and Singh, 1983). Though an array of pests attacks this crop, the major loss is inflicted by the pod borer, *M. vitrata* the larvae of which damage both vegetative and reproductive stages of the crop (Ganapathy, 2010). It is an important pest affecting the grain legumes in tropics and subtropics and is reported to feed on 39 host plants (Manjunath and Mallapur, 2015). At present, farmers rely mainly on synthetic chemical insecticides to overcome the problem of *M. vitrata*. Indiscriminate use of insecticide also leads to detrimental effect on natural enemies, environmental and potential health hazards for people working in the field.

Synergists are among the most straight forward tools for overcoming metabolic resistance because they can directly inhibit the resistance mechanism itself. Since the first demonstration of insecticide synergism over 79 years ago (Eagleson, 1940), their effective application against agricultural pests has offered tremendous promise but achieved little utility. This is partly because of difficulties in their use and partly because there was lack of basic understanding of insect detoxification systems.

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MATERIAL AND METHODS

Procedure for Bioassay

Each treatment consisted of ten larvae with four replications. Just before the insecticidal treatment, third instar larvae of *M. vitrata* were transferred carefully with camel brush in to clean and dry multi cavity cell trays at the rate of one larvae for one cell and covered with lid. The larvae were starved for two hours before treatment. For conducting bioassay studies flower buds dip bioassay method was used (Elzen *et al.*, 1992; Sreelakshmi *et al.*, 2015). 10 ml of quantity of five concentrations for each insecticide were prepared. Flower buds collected from the field were dipped in test insecticidal solutions for ten seconds and later shade dried. These flowers were placed in 9 cm diameter petriplate lined with filter paper; larvae were placed on these flowers and were allowed to feed on treated flowers. Control treatment was done with water alone.

Amount of insecticide required for preparation of required per cent concentration of insecticide solution was calculated by using below formula

Volume of insecticide required (ml) =

$$\frac{(\text{desired concentration of a.i in}\%) \times (\text{volume of stock solution in ml})}{(\% \text{ a.i in formulation})}$$

Data recorded

Larval mortality was taken 24 hour, 48 hours and 72 hours after treatment. The larvae which were moribund and did not show any movement were also considered as dead.

Statistical analysis

Data on mortality was subjected to Abbott's formula (Abbotts, 1925) for calculating corrected mortality percentage. LC₅₀ values were determined by probit analysis (Finney, 1971) using Statistical Packages for Social Sciences (IBM SPSS Statistics 20) software.

$$\text{Abbott's formula for corrected mortality} = \frac{T - C}{100 - C} \times 100$$

T : Mortality in treatment and

C : Mortality in control

After calculating the LC₅₀ value 24 HAT calculate the amount of a.i present in that concentration and mixing the 1:10 ratios (Lorini and Galley, 2000) of insecticide and synergist concentration, respectively. Synergists *viz.*, pyperonyl butoxide (PBO) and sesamin were used along with the test insecticides (at their respective LC₅₀ concentrations obtained from earlier bioassay experiment) against resistance larval population of *M. vitrata*, to determine the efficiency of synergists in enhancing the insecticide toxicity in the resistant larval population of *M. vitrata*. The insecticides *viz.*, chlorpyrifos, thiodicarb, spinosad, chlorantraniliprole in combination with PBO and dichlorvos in combination with sesamin in the ratios of 1:10 were evaluated by flowers dip method of bioassay insecticides and synergists were diluted with acetone to the required doses *i.e.*, 10 times more than the test doses of the insecticides (Table.1). The insecticides and synergists doses were mixed to get the insecticide synergist mixtures in 1:10 and test insect population were exposed till the mortality in the range of 5-90 per cent was recorded. Mortality was recorded 12 hours and 24 hours after treatment. Per cent mortality was calculated by using below formula

Per cent mortality =

$$\frac{\text{Number larvae dead}}{\text{Total number of larvae released}} \times 100$$

RESULTS AND DISCUSSION

The per cent larval mortality (12 HAT) in baseline population of *M. vitrata* was significantly higher when treated with chlorantraniliprole 67.78 per cent (significantly differs from other insecticide) followed by spinosad 61.62 per cent, thiodicarb 58.58 per cent (on par with each other), dichlorvos 52.53 per cent and chlorpyrifos 47.48 per cent (significantly differs from other insecticide) (Table 2).

The per cent larval mortality (24 HAT) in Baseline population of *M. vitrata* was significantly higher when treated with chlorantraniliprole 91.11 per cent followed by spinosad 88.89 per cent, thiodicarb 85.56 per cent, dichlorvos 83.33 per cent and chlorpyrifos 81.11 per cent (on par with each other) (Table 2).

There was significantly higher per cent mean larval mortality observed, when the larvae were treated with insecticide along with synergist (67.88%) as against

Table 1. Concentrations of synergist used for bioassay during *rabi*, 2018-2019

		Per cent concentration of LC₅₀ 24 HAT	Quantity of insecticide in µl or mg / 10 ml of water	Amount of a.i	Amount of Synergist µl
Chlorpyrifos	B.P	0.059	29.50	0.0059	0.059
	Prakasam	0.198	99.00	0.0198	0.198
	Nellore	0.218	109.00	0.0218	0.218
	Chittoor	0.291	145.50	0.0291	0.291
	Guntur	0.454	227.00	0.0454	0.454
	Kurnool	0.553	276.50	0.0553	0.553
Dichlorvos	B.P	0.074	9.74	0.007402	0.07402
	Prakasam	0.259	34.08	0.025901	0.25901
	Nellore	0.302	39.74	0.030202	0.30202
	Chittoor	0.384	50.53	0.038403	0.38403
	Guntur	0.509	66.97	0.050897	0.50897
	Kurnool	0.591	77.76	0.059098	0.59098
Thiodicarb	B.P	0.059	7.87	0.005903	0.05903
	Prakasam	0.179	23.87	0.017903	0.17903
	Nellore	0.199	26.53	0.019898	0.19898
	Chittoor	0.227	30.27	0.022703	0.22703
	Guntur	0.313	41.73	0.031298	0.31298
	Kurnool	0.371	49.47	0.037103	0.37103
Spinosad	B.P	0.082	18.22	0.008199	0.08199
	Prakasam	0.250	55.56	0.025002	0.25002
	Nellore	0.276	61.33	0.027599	0.27599
	Chittoor	0.348	77.33	0.034799	0.34799
	Guntur	0.402	89.33	0.040199	0.40199
	Kurnool	0.476	105.78	0.047601	0.47601
Chlorantraniliprole	B.P	0.002	1.08	0.0002	0.002
	Prakasam	0.008	4.32	0.000798	0.00798
	Nellore	0.009	4.86	0.000898	0.00898
	Chittoor	0.011	5.95	0.0011	0.011
	Guntur	0.013	7.03	0.001299	0.01299
	Kurnool	0.016	8.65	0.001599	0.01599

Insecticide resistance in larval population of *Maruca vitrata*

insecticide alone (47.27%) in 12 HAT and insecticide along with synergist (100%) as against insecticide alone (72.00%) in 24 HAT.

The per cent larval mortality (12 HAT) in Prakasam district population of *M. vitrata* was significantly higher when treated with chlorantraniliprole 59.60 per cent (significantly differs from other insecticides) followed by spinosad 52.53 per cent, thiodicarb 45.45 per cent (on par with each other), dichlorvos 42.42 per cent and chlorpyrifos 38.38 per cent (on par with each other) (Table 3).

The per cent larval mortality (24 HAT) in Prakasam district population of *M. vitrata* was significantly higher when treated with chlorantraniliprole 84.45 per cent (significantly differs from other insecticide) followed by spinosad 77.78 per cent, thiodicarb 74.45 per cent (on par with each other), dichlorvos 68.89 per cent and chlorpyrifos 64.45 per cent (on par with each other) (Table 3).

There was significantly higher per cent mean larval mortality observed, when the larvae were treated with insecticide along with synergist (56.97%) as against insecticide alone (38.88%) in 12 HAT and insecticide along with synergist (87.56%) as against insecticide alone (60.44%) in 24 HAT.

The per cent larval mortality (12 HAT) in Nellore district population of *M. vitrata* was significantly higher when treated with chlorantraniliprole 55.56 per cent (significantly differs from other insecticide) followed by spinosad 49.50 per cent (significantly differs from other insecticide), thiodicarb 43.44 per cent (significantly differs from other insecticide), dichlorvos 37.37 per cent and chlorpyrifos 34.34 per cent (on par with each other) (Table 4).

The per cent larval mortality (24 HAT) in Nellore district population of *M. vitrata* was significantly higher when treated with chlorantraniliprole 82.22 per cent (significantly differs from other insecticide) followed by spinosad 74.45 per cent, thiodicarb 71.11 per cent (on par with each other), dichlorvos 66.67 per cent and chlorpyrifos 62.22 per cent (on par with each other) (Table 4).

There was significantly higher per cent mean larval mortality observed, when the larvae were treated with insecticide along with synergist (52.93%) as against insecticide alone (35.15%) in 12 HAT and insecticide along with synergist (86.22%) as against insecticide alone (56.44%) in 24 HAT.

The per cent larval mortality (12 HAT) in Chittoor district population of *M. vitrata* was significantly higher when treated with chlorantraniliprole 46.47 per cent followed by spinosad 41.41 per cent, thiodicarb 35.35 per cent, dichlorvos 31.31 per cent and chlorpyrifos 28.28 per cent (on par with each other) (Table 5).

The per cent larval mortality (24 HAT) in Chittoor district population of *M. vitrata* was significantly higher when treated with chlorantraniliprole 75.56 per cent (significantly differs from other insecticide) followed by spinosad 68.89 per cent, thiodicarb 64.45 per cent (on par with each other), dichlorvos 58.89 per cent and chlorpyrifos 54.45 per cent (on par with each other) (Table 5).

There was significantly higher per cent mean larval mortality observed, when the larvae were treated with insecticide along with synergist (48.48%) as against insecticide alone (24.65%) in 12 HAT and insecticide along with synergist (77.78%) as against insecticide alone (51.11%) in 24 HAT.

The per cent larval mortality (12 HAT) in Kurnool district population of *M. vitrata* was significantly higher when treated with chlorantraniliprole 42.42 per cent (significantly differs from other insecticide) followed by spinosad 36.36 per cent (significantly differs from other insecticide), thiodicarb 31.31 per cent, dichlorvos 27.27 per cent and chlorpyrifos 25.25 per cent (on par with each other) (Table 6).

The per cent larval mortality (24 HAT) in Kurnool district population of *M. vitrata* was significantly higher when treated with chlorantraniliprole 70.00 per cent followed by spinosad 65.56 per cent (significantly differs from other insecticide), thiodicarb 57.78 per cent (on par with each other), dichlorvos 53.33 per cent and chlorpyrifos 48.89 per cent (on par with each other) (Table 6).

There was significantly higher per cent mean larval mortality observed, when the larvae were treated with insecticide along with synergist (42.83%) as against insecticide alone (22.22%) in 12 HAT and insecticide along with synergist (71.56%) as against insecticide alone (46.67%) in 24 HAT.

The per cent larval mortality (12 HAT) in Guntur district population of *M. vitrata* was significantly higher when treated with chlorantraniliprole 37.37 per cent (significantly differs from other insecticide) followed by spinosad 31.31 per cent (significantly differs from other

Table 2. Effect of synergism on the insecticide toxicity of baseline larval population of *M. virata* during *rabi*, 2018-19

Insecticide	12 HAT			24 HAT		
	Without Synergist	With Synergist	Mean	Without Synergist	With Synergist	Mean
Chlorpyrifos 20% EC	38.38 (31.84)	56.57 (40.66)	47.48 ^a	62.22 (43.33)	100.00 (75.00)	81.11 ^a
Dichlorvos 76% EC	40.40 (32.81)	64.65 (44.69)	52.53 ^b	66.67 (45.71)	100.00 (75.00)	83.33 ^a
Thiodicarb 75% WP	46.46 (35.80)	70.71 (47.92)	58.58 ^c	71.11 (48.34)	100.00 (75.00)	85.56 ^a
Spinosad 45% SC	52.53 (38.72)	70.71 (47.92)	61.62 ^c	77.78 (51.81)	100.00 (75.00)	88.89 ^a
Chlorantraniliprole 18.5% SC	58.59 (41.68)	76.77 (51.09)	67.68 ^d	82.22 (54.70)	100.00 (75.00)	91.11 ^a
Mean	47.27 ^a	67.88 ^b	57.58	72.00 ^a	100.00 ^b	86.00
LSD @ 0.05		7.56	4.78	5.88	5.88	3.72

*Values in parenthesis are angular transformed values

Table 3. Effect of synergism on the insecticide toxicity of *M. virata* larval population collected from Prakasam district during *rabi*, 2018-19

Insecticide	12 HAT			24 HAT		
	Without Synergist	With Synergist	Mean	Without Synergist	With Synergist	Mean
Chlorpyrifos 20% EC	28.28 (26.61)	48.48 (36.77)	38.38 ^a	48.89 (36.94)	80.00 (53.00)	64.45 ^a
Dichlorvos 76% EC	34.34 (29.86)	50.51 (37.74)	42.42 ^a	53.33 (39.12)	84.44 (55.89)	68.89 ^a
Thiodicarb 75% WP	38.38 (31.84)	52.53 (38.72)	45.45 ^{ab}	60.00 (42.34)	88.89 (60.58)	74.45 ^b
Spinosad 45% SC	40.40 (32.70)	64.65 (44.80)	52.53 ^{ab}	66.67 (46.19)	88.89 (60.58)	77.78 ^b
Chlorantraniliprole 18.5% SC	50.51 (37.75)	68.69 (46.79)	59.60 ^b	73.33 (49.53)	95.56 (68.51)	84.45 ^c
Mean	38.38 ^a	56.97 ^b	47.68	60.44 ^a	87.56 ^b	74.00
LSD @ 0.05		8.25	5.22	8.67	8.67	5.48

*Values in parenthesis are angular transformed values

Table 4. Effect of synergism on the insecticide toxicity of *M. vitrata* larval population collected from Nellore district during rabi, 2018-19

Insecticide	12 HAT			24 HAT		
	Without Synergist	With Synergist	Mean	Without Synergist	With Synergist	Mean
Chlorpyrifos 20% EC	24.24 (24.14)	44.44 (34.79)	34.34 ^a	44.44 (34.81)	80.00 (53.26)	62.22 ^a
Dichlorvos 76% EC	28.28 (26.61)	46.46 (35.76)	37.37 ^a	48.89 (36.97)	84.44 (55.89)	66.67 ^a
Thiodicarb 75% WP	36.36 (30.87)	50.51 (37.74)	43.44 ^b	55.56 (40.21)	86.67 (57.33)	71.11 ^b
Spinosad 45% SC	40.40 (32.85)	58.59 (41.68)	49.50 ^c	62.22 (43.53)	86.67 (57.33)	74.45 ^b
Chlorantraniliprole 18.5% SC	46.46 (35.76)	64.65 (44.69)	55.56 ^d	71.11 (48.09)	93.33 (65.26)	82.22 ^c
Mean	35.15 ^a	52.93 ^b	44.04	56.44 ^a	86.22 ^b	71.33
LSD @ 0.05	8.30			8.19		

*Values in parenthesis are angular transformed values

Table 5. Effect of synergism on the insecticide toxicity of *M. vitrata* larval population collected from Chittoor district during rabi, 2018-19

Insecticide	12 HAT			24 HAT		
	Without Synergist	With Synergist	Mean	Without Synergist	With Synergist	Mean
Chlorpyrifos 20% EC	18.18 (20.65)	38.38 (31.84)	28.28 ^a	40.00 (32.66)	68.89 (47.25)	54.45 ^a
Dichlorvos 76% EC	20.20 (21.76)	42.42 (33.82)	31.31 ^a	44.44 (34.79)	73.33 (49.53)	58.89 ^a
Thiodicarb 75% WP	24.24 (24.49)	46.46 (35.76)	35.35 ^a	51.11 (38.03)	77.78 (51.81)	64.45 ^b
Spinosad 45% SC	28.28 (26.61)	54.55 (39.69)	41.41 ^a	55.56 (40.19)	82.22 (54.45)	68.89 ^b
Chlorantraniliprole 18.5% SC	32.32 (28.74)	60.61 (42.69)	46.47 ^a	64.44 (44.75)	86.67 (57.33)	75.56 ^c
Mean	24.65 ^a	48.48 ^b	36.57	51.11 ^a	77.78 ^b	64.45
LSD @ 0.05	8.145			9.46		

*Values in parenthesis are angular transformed values

Insecticide	12 HAT			24 HAT		
	Without Synergist	With Synergist	Mean	Without Synergist	With Synergist	Mean
Chlorpyrifos 20% EC	16.16 (19.53)	34.34 (29.86)	25.25 ^a	37.78 (31.57)	60.00 (42.47)	48.89 ^a
Dichlorvos 76% EC	18.18 (2065)	36.36 (30.87)	27.27 ^a	40.00 (32.63)	66.67 (46.19)	53.33 ^a
Thiodicarb 75% WP	20.20 (22.01)	42.42 (33.82)	31.31 ^a	44.44 (34.81)	71.11 (48.34)	57.78 ^{ab}
Spinosad 45% SC	24.24 (24.49)	48.48 (36.77)	36.36 ^b	53.33 (39.12)	77.78 (51.81)	65.56 ^b
Chlorantraniliprole 18.5% SC	32.32 (28.74)	52.53 (38.72)	42.42 ^c	57.78 (41.28)	82.22 (54.45)	70.00 ^b
Mean	22.22 ^a	42.83 ^b	32.53	46.67 ^a	71.56 ^b	59.12
LSD @ 0.05		6.61#	4.18	8.37	8.37	5.30

*Values in parenthesis are angular transformed values

Insecticide	12 HAT			24 HAT		
	Without Synergist	With Synergist	Mean	Without Synergist	With Synergist	Mean
Chlorpyrifos 20% EC	12.12 (16.81)	28.28 (26.61)	20.20 ^a	33.33 (29.29)	46.67 (35.88)	40.00 ^a
Dichlorvos 76% EC	14.14 (17.92)	32.32 (28.74)	23.33 ^a	35.56 (30.35)	53.33 (39.12)	44.45 ^a
Thiodicarb 75% WP	16.16 (19.53)	36.36 (30.83)	26.26 ^{ab}	40.00 (32.66)	60.00 (42.47)	50.00 ^b
Spinosad 45% SC	20.20 (21.76)	42.42 (33.82)	31.31 ^b	44.44 (34.81)	75.56 (50.62)	60.00 ^b
Chlorantraniliprole 18.5% SC	26.26 (25.60)	48.48 (36.77)	37.37 ^c	48.89 (36.94)	77.78 (51.81)	63.34 ^b
Mean	17.78 ^a	37.58 ^b	27.68	40.44 ^a	62.67 ^b	51.56
LSD @ 0.05		7.22	4.56	8.55	8.55	5.41

*Values in parenthesis are angular transformed values

insecticide), thiodicarb 26.26 per cent, dichlorvos 23.33 per cent and chlorpyrifos 20.20 per cent (on par with each other) (Table 7).

The per cent larval mortality (24 HAT) in Guntur district population of *M. vitrata* was significantly higher when treated with chlorantraniliprole 63.34 per cent followed by spinosad 60.00 per cent, thiodicarb 50.00 per cent (on par with each other), dichlorvos 44.45 per cent and chlorpyrifos 40.00 per cent (on par with each other) (Table 7).

There was significantly higher per cent mean larval mortality observed, when the larvae were treated with insecticide along with synergist (37.58%) as against insecticide alone (17.78%) in 12 HAT and insecticide along with synergist (62.67%) as against insecticide alone (40.44%) in 24 HAT.

In present investigations more per cent larval mortality was observed in insecticide + synergist treated larval population when compared with insecticide alone treated population. These findings were supported by Yu (1983) who revealed that piperonyl butoxide, the well-known inhibitor of microsomal oxidases, enhanced the toxicity of OP compound insecticides in *M. vitrata* and also Armes *et al.* (1997) reported that treatment with the metabolic inhibitor, piperonyl butoxide, resulted in complete suppression of cypermethrin resistance (2- to 121-fold synergism), indicating that enhanced detoxification by microsomal P450-dependent monooxygenases was probably the major mechanism of pyrethroid resistance. Wang *et al.* (2009) reported that after inhibitors were used, spinosad resistance could be partially suppressed by piperonylbutoxide (PBO) and triphenylphosphate (TPP), but not by diethyl maleate (DEM) in *Helicoverpa armigera*. Bao *et al.* (2014) reported that the synergist, piperonyl butoxide, respectively caused 1.1-fold, 5.8-fold and 9.0-fold decreases in the resistance ratios of the OK, TS1, and TS5 strains of *Thrips palmi* against spinosad. The maximum per cent mortality reached 48 HAT because of these synergists were inhibit the mechanism of detoxification enzymes in larval population of *M. vitrata*. Present results were also supported by results of Wang *et al.* (2018) who conducted chlorpyrifos synergist (PBO) experiment and indicated that the Lab-HN strain and field-collected population of *S. litura* from Shuangliu (China) in 2016 (SL16 population) showed that, the highest synergism ratios (SR) of 34.4 and 73.3 fold respectively.

The results of Bingham *et al.* (2008) were also in agreement with present findings, stated that PBO reduced the pirimicarb resistance factor in a clone of *Myzus persicae* (Sulzer) from >19000 to 100 fold and in *Aphis gossypii* (Glover) from >48000 to 30 fold. Similar results were obtained for a strain of *Bemisia tabaci* Gennadius. Synergism was also observed in laboratory susceptible insects, suggesting that, even when detoxification is not enhanced, there is degradation of insecticides by the background enzymes. Use of an analogue of PBO, which inhibits esterases but has reduced potency against microsomal oxidases, suggests that acetamiprid resistance in whiteflies is largely oxidase based. Present results are also supported by Sohail and Irfanullah (2004) who used sesame oil, as the synergist. The insecticide was mixed with sesame oil in 1:1 and 1:2 ratios to obtain LC₅₀ of the insecticide + synergist combination. House fly was allowed to feed on the insecticide coated sugar for 48 hr. The mortality data was observed after 1, 2, 4, 6, 8, 12, 24 and 48 hours. The results showed that dichlorvos was the most effective with LC₅₀ value, 122 ppm and with the synergist combination.

Role of synergist for countering the insecticide resistance mechanism in all populations were studied and the highest percent mortality was observed in larval population treated with chlorantraniliprole alone and along with synergist followed by spinosad, thiodicarb, dichlorvos and chlorpyrifos alone and along with synergist respectively. Chlorantranilip -role in combination with piperonyl butoxide gave maximum mortality in all population of *M. vitrata* 12 and 24 HAT. Maximum per cent mortality was observed in baseline population followed by Prakasam, Nellore, Chittoor, Kurnool and Guntur during *rabi* 2018-19.

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